A Recent Update on Hepatoprotective Potential of Herbal Plant

Mohammad Irfan Ali¹ and Manoj Kumar²

¹School of Sciences, Suresh Gyan Vihar University, Jaipur-302017, Rajasthan, India

²Marine Biotechnology Laboratory, Department of Botany, University of Delhi, North Campus, India

*Corresponding Author- e-mail: irfan_biochem07@gmail.com

Abstracts

Liver damage is an epidemic and metabolic disorder which is the most important cause of mortality and morbidity globally. Liver injury treatments are important issue of today's research domain, because of many allopathic drugs and their toxic influence lead to liver damage. Therefore attention is drawn to the potentials of medicinal plants that have the hepatoprotective ability to reduce or cure liver disorder. The use of herbal plants or their primary and secondary metabolites for curing diseases has long being in continuation since ancient times due to its therapeutic efficacy and safety. Various herbal plants have been investigated for their hepatoprotective potential to treat different types of liver disorder. Numerous herbal plants and formulations are effective in treatment of liver disorder. This systemic review mainly is focused on herbal plants as hepatoprotective in various traditional medicines and explores the herbal plant, isolated active constituent and formulation with hepatoprotective activity.

Keywords: Hepatoprotective activity; Herbal plant; Liver disease

Introduction

Liver is the largest glandular organ of the body which plays a pivotal role to regulate whole metabolic process and homeostasis of the body. It is the most important site of intermediary metabolism and accountable for detoxifying any foreign material and other xenobiotics by converting and excreting waste and toxin. It is considered as one of the most vital organs due to the handling metabolism of carbohydrates, lipid, protein, secretion of bile, storage of vitamins and production of a variety of coagulation factors. Thus the maintenance of healthy liver is imperative for human health (Pradhan and Girish 2006, Haidry and Malik 2014). In spite extensive regenerative

continuous various exposures to environmental pollutants, xenobiotics, and chemotherapeutic agents could repress and overcome the natural protective ability of the liver, leading to liver malfunction and later if it is not treated properly leads to liver injury.

Today, alcohol misuse is one of the major health problems globally. There is a close relationship between ethanol intake and alcoholic liver disease because of 80% of consumed alcohol is metabolized in the liver consequently profound effect on the metabolism of lipids lipoproteins. and Moreover, ethanol is metabolized into cytotoxic acetaldehyde by enzyme alcohol dehydrogenase and acetaldehyde is oxidized to acetate by aldehyde oxidase or xanthine

oxidase in the liver, giving rise to reactive oxygen species (ROS) via cytochrome P450 2E1 (Lu and Cederbaum 2008). This leads to oxidative stress in the hepatic cells which is the most striking initial expression of alcoholinduced liver injury ultimately damage to the liver cell membrane and the cytosolic enzymes are leaked into the blood stream. Therefore, the elevation of these cytosolic enzymes in the blood stream serves as a quantitative marker of hepatic damage (Ramaiah 2007).

It has been reported that the free radicals induced oxidative stress is a main cause of liver disorder such as swelling, necrosis, degeneration and apoptosis of hepatic cells. Liver may be damaged by free radicals via mechanisms of lipid peroxidation and covalent binding with consequent tissue injury. Reactive oxygen species (ROS) such as superoxide, hydroxyl, peroxyl, and alkoxy radicals cause damage to nucleic acid, proteins, and membrane lipids and have been associated with many aging related problems together with atherosclerosis, liver disorders, lung, kidney damage, diabetes mellitus, cancer inflammatory diseases and cardiovascular diseases (Singh et al. 2008, Pal et al. 2014). Lipid peroxidation damage to cell membranes subsequent affects the integrity and function of the cell membrane which affects the cell's ability to maintain ion gradients and transport. On the other hand liver disorders are mainly caused by chemicals and drugs when taken in very high doses (Ekaidem et al. 2012).

In case of liver damages the capacity of natural antioxidant system is impaired. ROS are

generated by environmental factors such as ultraviolet radiation, pollutants, x-rays, or by normal metabolic process in mitochondria. The intracellular concentration of ROS depends on both their generation by endogenous or exogenous factors and removal by various endogenous antioxidants including both enzymatic and nonenzymatic processes (Haque et al. 2014)

Liver diseases are major global health problem prevalent in developing countries. Liver disease classified as hepatosis (noninflammatory), acute or chronic hepatitis (inflammatory) and cirrhosis or fibrosis (degenerative). It is frequently abused by the environmental toxins, heavy metals, poor eating habits, alcohol, prescription and the counter drug use thus it is damaged and weakened ultimately leads to the hepatitis, jaundice, liver fibrosis and alcoholic liver disease. Liver disease may results in elevated levels of plasma total cholesterol, Low density lipoprotein cholesterol (LDL-C), and Triacylglycerols (TGs) are associated with high risk of atherosclerosis and cardiovascular disease (Dominiczak, 2005, Ekaidem et al. 2012).

Almost all organisms possess antioxidant defence and repair systems which reduce the production of free radical species, but these protective systems are insufficient to entirely prevent the damage when there is increased oxidant radical generation. Various antioxidant agents of herbal plant have been found to protect human liver from free radicals damage (Ekaidem et al. 2012). The presence of a wide

range of phytoconstituents such as phenolics thiols and caretonoids in herbal plants protects the human body against oxidative damage by ROS (Pal et al. 2014).

Although the notable in progresses conventional medical therapy in the last 20 years, drugs available for the treatment of hepatic diseases were limited in efficacy and could have prompted various unwanted side effects when compared to other medical therapies for hepatic diseases which were difficult to handle. In addition, some of these modern hepatoprotective drugs did not protect liver against injury. In response to these factors that limit the use of conventional drugs and efforts were continuously made to identify new sources of agents with hepatoprotective potential (Mamat et al. 2013).

Interestingly, over the last few decades the reputation use complementary of alternative medicines has increased worldwide due to its therapeutic efficiency and safety, particularly herbal/plant-based therapies, to cure various diseases, efforts are increasingly being carried out by scientists to investigate the hepatoprotective potential of various medicinal plants (Mamat et al. 2013). The plant kingdom is a valuable source of new herbal medicinal agents. In recent years numerous traditional medicinal plants were tested for their hepatoprotective potential in the experimental animals. Traditional medicines and prescriptions with beneficial against various pathological conditions have recently paying attention as alternative therapies (Biswas et al. 2014). Several studies

have proved the beneficial outcomes of herbal medicine for human health. A variety of molecules have been isolated and their physicochemical and pharmacological properties have been studied.

However, compounds and extracts need to be appropriately formulated to facilitate their physiological target and pharmacological activity. Factors such as low permeability and solubility could affect the absorption and delivery of bioactive molecules (Fang and Bhandari, 2011). On the other hand the shelflife of herbal medicine should be evaluated in order to assurance the stability during the period of use. Degradation reactions are enhanced by temperature, humidity, pH, oxygen and light. Herbal medicines are complex mixtures of different classes of chemical compounds, such as carbohydrates, lipids, proteins, and secondary metabolites (Bott et al. 2010).

In present study an effort has been made to review the most prospective herbal plants having pharmacologically most reputable hepatoprotective potential.

Herbal plant with reported hepatoprotective potential

Allium cepa

Allium cepa (Family: Liliaceae) commonly known as garden onion. A. cepa is a bulbous plant extensively cultivated in China, India and United States. It is rich in carbohydrates, potassium, sodium and phosphorus. Traditionally onion has been used to treat

intestinal infections, ear ache, eye infections, headaches associated with drowsiness, urinary tract burning, ulcers on heels and cough resulted from inspiration of cold air (Price et al. 1997).

Phytochemical screening revealed that aqueous extract of bulbs of A. cepa contains abundant a group of polyphenolic compounds tannins, saponins, flavanoids. organosulphur compounds like dipropyl disulphide, methyl-1propenyl trisulphide and propyl-1-propenyl trisulphide (Teyssier et al. 2001). Approximately 20 types of flavonols were detected, with the two main flavonols representing up to 80–85% of the total flavonoid content i.e. quercetin conjugates:quercetin-m 3, 40-O-diglucoside (QDG) and quercetin-40-O-monoglucoside (QMG) (Price et al. 1997).

Many reports revealed that onion was found to have antibacterial, antiviral, antiparasitic, antihyperlipidemic, antifungal, antiinflammatory antihypertensive, antihypoglycemic, antithrombotic, and antioxidant activities (Griffiths et al. 2002). In recent decades, the extracts of various parts of the A. cepa have been extensively studied for their anti-diabetic (Kook et al. 2009), anti-tumor (Dorant et al. 1996), hepatoprotective (Obioha et al. 2009), and anti-nephrotoxicity (Ige et al. 2011) activities. The antioxidant activity in A. cepa bulb is mainly due to the existence of flavonoids, selenium, vitamin C and amino acids (Zielinska et al. 2003, Elhassaneen et al. 2009). A. cepa also shows effective antioxidant activity.

A. cepa aqueous bulb extract had showed hepatoprotective activity against ethanolinduced hepatotoxicity in adult male albino wistar rats which might be due to its antioxidant potential against DPPH (2,2diphenyl-1-picrylhydrazyl), hydroxyl and superoxide radicals which quench ROS and regenerate membrane-bound antioxidants at both preventive and curative doses and significantly restored the elevated levels of Serum aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP) and total bilirubin (TB) (Kumar et al. 2013).

Alocasia indica

Alocasia indica (Family: Araceae) extensively cultivated in tropical and subtropical regions, such as West Bengal, Maharashtra Assam, and Southern India. In local languages of West Bengal, it is popular as 'Mannkochu'. The plant is a perennial herb frequently attaining heights of 5 meters. The tuber part of the A. indica plant is edible and is also used as a common vegetable available easily and cheap among common people (Pal et al. 2014). Since last few decades A. indica is used in the treatment of abdomen and spleen related disorders. The non-edible part of the plant, like leaves, has been used for the treatment of various inflammatory diseases including bruises and rheumatism (Pal et al. 2014).

It was also revealed that the ehanolic extract of the leaves of the plant contained free radical scavenging activity, hepatoprotective activities (Mulla et al. 2009), antioxidant, antinociceptive, anti-inflammatory, antimicrobial activities (Mulla et al 2010), antidiarrheal and antiprotozoal activities (Mulla et al. 2011).

Phytochemical screening of both the ehanolic and aqueous extract revealed the presence of alkaloids, flavonoids, tannins, glycosides and phenolics as found in several plant extract (Vulgaris et al. 2013).

It has been reported that phenolic compounds like flavonoids, phenolic acid, proanthocyanidins, diterpenes and tannins are derived from several medicinal plants and exhibit antioxidant activity by inactivating free radicals or by preventing the decomposition of hydroperoxides into free radicals. Flavonoids have free radical scavenging activity, inhibition of hydrolytic and oxidative enzyme activity as well as antinociceptive and antiinflammatory activities (Akinpelu et al 2010, Pal et al. 2014).

In vivo antioxidant activity was done by hepatoprotection against liver injury induced by carbon tetrachloride (CCl₄) in rats which has been extensively and successfully reported by many investigators. CCl₄ is metabolized by cytochrome P-450 in endoplasmic reticulum and mitochondria with the formation of CCl₃O , a ROS which initiates lipid peroxidation and enhanced marker enzymes such as ALT, ALP and AST (Shahjahan et al. 2004). Malonaldehyde (MDA), marker of lipid peroxidation, was noticeably increased in ethanol-attenuated liver. Reduced glutathione

(GSH) is normally present in hepatocytes for detoxification of free radicals. GSH depletion increases the sensitivity of cells to various aggressions leading to tissue disorder and injury (Meagher et al. 1999, Pal et al. 2014).

Histopathological study of CCl₄ induced liver section shows severe necrotic changes and substantial changes such as microvesicular steatosis, increase in sinusoidal space, inflammatory infiltration of lymphocytes, dilation of central vein and increase in fat droplet (Shi et al 1998).

Ethanolic extract of *A. indica* tuber recover the changes by showing the normal pattern of the central vein, radiating pattern of cell plates and absence of fat droplets. Serum marker ALT, ALP, AST, MDA, GSH were significantly reversed by ethanolic extract of *A. indica* tuber (Pal et al. 2014).

Antrodia Cinnamomea

Antrodia cinnamomea (Family: Polyporaceae) is a medicinal mushrooms and parasitic fungus can be mostly found in Taiwan. A. Cinnamomea is used to treat liver diseases, abdominal pain, hypertension, diarrhea, and other diseases (Ao et al. 2009). Historically, indigenous Taiwanese people have used to treat liver disorders from excessive alcohol consumption. However, it is difficult to collect wild A. cinnamomea fruiting bodies (Liu et al. 2012).

Several reports have been published on the chemical components of *A. cinnamomea*. Phytochemical constituents isolated from *A.*

cinnamomea, such as flavonoids, benzenoids, polyphenols, maleic/succinic acid derivatives, diterpenes, triterpenoids, steroids, and polysaccharides, have been reported to exhibit therapeutic activities (Lu et al. 2007 and 2011, Huang et al. 2010a, Kumar et al. 2011, Wu et al. 2011).

A. Cinnamomea fruiting bodies were reported to show the hepatoprotective potential against ethanol induced acute liver injury in Sprague Dawley (SD) rats (Chiang et al., 2010). Huang et al. investigated the effects of A. cinnamomea fruiting bodies against chronic consumption in rats and found that the gene expression of SREBP-1c, acetyl-CoA 3-hydroxy-3-methoxyglutarylcarboxylase, CoA reductase, fatty acid synthase and malic enzyme was downward regulated. The results of histopathological examination also revealed that the alcohol induced liver injuries i.e. hepatocyte necrosis and inflammatory cell infiltration, were prevented by ethanolic extract A. cinnamomea fruiting body (Huang et al. 2010a).

Wu et al. investigated the hepatoprotective potential of *A. cinnamomea* fruiting bodies against ethanol induced liver fibrosis and found that the expression of hepatic mRNAs, i.e., matrix metalloproteinase (MMP)-9, tumor necrosis factor (TNF)-, Kruppel-like factor (KLF)-6 and transforming growth factor (TGF)-1, were downward regulated by ethanolic extract *A. cinnamomea* (Wu et al. 2011).

Lu et al. reported the effects of *A. cinnamomea* mycelia against ethanol induced liver injury in male SD rats and found the suppression of elevated levels of AST, ALT, ALP, and TB in rats by orally administration of *A. cinnamomea*. Lu et al. (2011) further reported that a triterpenoid enriched fraction of the ethanolic extract of *A. cinnamomea* mycelia showed the greatest effectiveness in preventing ethanol induced acute liver injury and free radical generation in rats (Lu et al. 2007).

Hsiao et al. examined the hepatoprotective potential of the water extract of fruiting bodies against CCl4 induced liver injury in mice and found that the level of plasma transaminases, i.e., AST and ALT, were significantly decline in the group of mice orally administered with the aqueous extract of A. cinnamomea (Hsiao et al. 2003). Hseu et al., reported that the activities of hepatic superoxide dismutase (SOD) and catalase (CAT) were enhanced by treatment with the water extract of A. cinnamomea. The water extract of A. cinnamomea fruiting bodies exhibits an ability 1,1-diphenyl-2-picrylhydrazyl scavenge radicals in an in vitro study (Hseu et al., 2008).

Bidens pilosa

Bidens. pilosa (Family: Asteraceae) is an erect, perennial herb widely distributed from temperate and tropical regions (Alcaraz et al. 1998) and traditionally medicine used for several purposes including the treatment of antitumor, anti-inflammatory, antidiabetic and antihyperglycemic, antioxidant, immunomodulatory, antimalarial, antibacterial,

antifungal, antihypertensive, vasodilatory, hepatoprotective and antiulcerative activities. (Yuan et al. 2008, Arlene et al. 2013).

B. pilosa is an extraordinary source of phytochemicals and 201 compounds have so far been identified from this plant, including 70 aliphatics (36 polyynes), 60 flavonoids, 25 terpenoids, 19 phenylpropanoids, 13 aromatics, 8 porphyrins, and 6 other compounds (Silva et al 2011).

All parts of the plant are used in the medicinal preparations, which are mainly water and alcohol macerations in bottles or herbal teas used orally or topically. Kviecinski et al. evaluated free radical scavenging activity of *B. pilosa*, which exert a beneficial action in preventing liver damage induced by CCl₄ and significant decrease serum enzymatic activities of AST, ALT, and lactate dehydrogenase (LDH) (Kviecinski et al. 2011).

Boerhavia diffusa

Boerhavia diffusa (Family: Nyctaginaceae) is a well known medicinal plant in traditional Indian medicine as well as Southern American and African continent. Its various parts and especially roots have been used for gastrointestinal, hepatoprotective, and gynecological indications, immunomodulation, antifibrinolysis, anticancer activity, antidiabetic activity, anti-inflammation, and diuresis. B. diffusa has been widely studied for its phytochemical constituents and therapeutic activities. The roots are the source of a novel class of isoflavonoids known as rotenoids,

flavonoids, flavonoid glycosides, xanthones, purine nucleoside, lignans, ecdysteroids, and steroids (Mishra et al. 2014a).

Miralles et al. reported 15 amino acids (6 essential) in the whole plant and 14 amino acids (7 essential) in the roots along with isopalmitate acetate, behenic acid, arachidic acid (6.3%), and saturated fatty acids (38%) (Miralles et al. 1988). Ujowundu et al. (2008) accounted the presence of vitamins C, B3, and B2 (44.80, 97.00mg, and 22.00mg) along with calcium (174.09mg) in roots. B. diffusa contains various categories of secondary metabolites, i.e. flavonoid glycosides, isoflavonoids (rotenoids), steroids (ecdysteroid), alkaloids, and phenolic and lignin glycosides (Bairwa et al. 2014).

Venkatalakshmi et al. accounted for protection against paracetamol induced hepatotoxicity for B. diffusa extracts (Venkatalakshmi, 2011). Olaleye et al. evaluated the aqueous and ethanolic extracts of fresh leaves for antioxidant activity in vitro and in vivo assays. Antioxidative evaluation of the ethanolic extract has shown appreciable quantities of phenolic and flavonoid content along with vitamins C and E. It also contained selenium and zinc. Pretreatment with B. diffusa aqueous and ethanolic extracts decline enzymatic activities and serum bilirubin caused by acetaminophen. The increase in ALP was reduced by almost 50% by aqueous and ethanolic extracts (both 400mg/Kg, orally for 7 days) whereas the increase in ALT and AST was decreased by more than 70% and serum

LDH level was also restored (Olaleye et al. 2010).

Caesalpenia crista

Caesalpinia Febaceae), crista (Family: commonly known as karanja, is an extensive shrub perennial climber distributed throughout India on waste land and coastal areas (Gupta et 2005). Phytochemical screening of ethanolic extract of C. crista revealed the presence of various phyto-constituents such as alkaloids, saponin glycosides, phenolic, flavonoids and carbohydrate. The seeds of the plant contain arginine, aspartic acid, citroline and diterpine -caesalpin.

C. crista have been used for treatment of antiinflammatory, jaundice, antihelmintic, antimalarial, stomachic, various liver disorders antidiabeti and antiperiodic, antipyretic (Kannur et al. 2006).

The ethanolic extract of *C. crista* (200 and 400 mg/kg) significantly reversed the levels of serum glutamic pyruvates transaminase (SGPT) and serum glutamic oxaloacetate transaminase (SGOT) and ALP and total bilirubin and triglycerides against paracetamol induced liver toxicity in rats.

Chelidonium majus

Chelidonium majus L. (family: Papaveraceae) is a plant highly admire for its therapeutic potential in western phytotherapy and traditional Chinese medicine (TCM). The plant contains, as major phytochemicals i.e. isoquinoline, flavonoids, phenolic acids and

alkaloids such as sanguinarine, chelidonine, chelerythrine, berberine, protopine and coptisine. Both crude extracts of *C. majus* and purified compounds derived from it exhibit a wide variety of biological activities i.e. anti-inflammatory, antimicrobial, immunomodulatory, antitumoral, choleretic, hepatoprotective, analgesic (Gilcaa et al. 2010).

The ethanolic extract of whole plant exerted marked hepatoprotection against CCl₄ toxicity in rats and p-dimethylaminoazobenzene-induced hepatocarcinogenesis in mice, indicated by a reduction in the number of necrotic cells, a prevention of fibrotic changes, and decreased activities of transaminases and bilirubin (Biswas et al. 2008).

Cyperus rotundus

Cyperus rotundus L. (Family: Cyperaceae), also known as purple nutsedge or nutgrass, is a common perennial weed with slender, scaly creeping rhizomes, bulbous at the base and arising singly from the tubers which are about 1-3 cm long (Uddin et al. 2006).

Different phytochemical studies on *C.rotundus* revealed the presence of alkaloids, flavonoids, tannins, starch, glycosides, furochromones, monoterpenes, sesquiterpenes, sitosterol, glycerol, linolenic, myristic and stearic acids. The major phytochemicals isolated from essential oil and the extracts of *C. rotundus* rhizome are Alpha-cyperone, Alpha-rotunol, Beta-cyperone, Beta-pinene, Beta-rotunol, Beta-selinene, Calcium, Camphene, Copaene,

Cyperene, Cyperenone, Cyperol, Cyperolone Cyperotundone D-copadiene, D-epoxyguaiene, D-fructose, D-glucose, Flavonoids, Gammacymene, Isocyperol, Isokobusone, Kobusone, Limonene. Linoleic-acid, Linolenic-acid, Magnesium, Manganese, C. rotunduskone, Myristic-acid, Oleanolic-acid, Oleanolic-acid-3-o-neohesperidoside, Oleic-acid, P-cymol, Patchoulenone. Pectin. Polyphenols, Rotundene. Rotundenol. Rotundone, Selinatriene, Sitosterol, Stearic-acid, Sugeonol, Sugetriol [Sivapalan 2013].

C.rotundus contains an essential oil that provides for the characteristic odour and taste of the herb, comprised mostly sesquiterpene hydrocarbons, epoxides, ketones. monoterpenes and aliphatic alcohols. Sesquiterpenes include selinene, isocurcumenol, nootkatone, aristolone, isorotundene. cypera-2,4(15)-diene, and norrotundene, as well as the sesquiterpene alkaloids rotundines A-C. Other constituents include the ketone cyperadione, and the monoterpenes cineole. camphene limonene. C. rotundus has also been shown to contain miscellaneous triterpenes including oleanolic acid and sitosterol, as well as flavonoids, sugars and minerals (Jeong et al. 2000, Kilani et al. 2008).

A number of pharmacological and biological activities including anti-inflammatory, antidiabetic, antidiarrhoeal, cytoprotective, antimutagenic, antimicrobial, antibacterial, antioxidant, cytotoxic, apoptotic, anti-pyretic and analgesic activities have been reported for this plant [Sivapalan 2013].

The ether extract of the rhizomes of *C. rotundus* (100 mg/kg) exhibited significant hepatoprotective effect by lowering serum levels of glutamic oxaloacetic transaminase, glutamic pyruvic transaminase, alkaline phosphatase and total bilirubin in rats by inducing liver damage by carbon tetrachloride. The ethyl acetate extract (100 mg/kg) exhibited hepatoprotective activity supplemented by histopathological examination of liver sections [Kumar et al. 2005, Sivapalan 2013].

The methanolic extract of leaves of *c. rotundus* (200mg/kg) evaluated for the hepatoprotective effect against CCl₄ induced liver damage in wistar albino rats which shows significant (p<0.05) hepatoprotective effect by lowering the SGOT, SGPT and ALP (Rao et al. 2014).

Dendrophthoe falcate

Dendrophthoe falcate (Family: Loranthaceae) is used extensively in traditional system of medicine as cooling, bitter, aphrodisiac, epilepsy, astringent, narcotic, diuretic, pulmonary tuberculosis, asthma, menstrual disorders, swellings, wounds, ulcers, renal, vesical calculi and skin diseases. The decoction of whole plant is used to treat joint pains and leaf juice is used for relief from chest pain. D. falcate is also reported to have cytotoxic, immunomodulatory activities, and wound healing potentials (Haque et al 2014).

Leaves of *D. falcate* contain L- Threonine dehydratase, hexokinase, Glucanphosphatase. It has also been reported several possible active chemical constituents such as -

amyrinacetate, -sitostirol, stigmasterol, oleanolic acid, kaempferol, quercetin, quercetin-3-O-rhamnoside, rutin, myricetin and their glycosides, catechin, leucocyanidin, kaempferol-3-O- -L-rhamnopyranoside quercetin-3-O- -Lrhamnopyranoside. D. falcate also contains tannins comprising of gallic acid, chebulinic acid, ellagicacid, quercetinand and cardiac glycosides such as strospeside, odoroside F and neritaloside were isolated from the leaves of D. falcate. Pentacyclic triterpenes: 3 -acetoxy-1 -(2hydroxy-2-propoxy)-11 -hydroxy-olean-12ene, kaempferol-3-O- -L-rhamnopyranoside, quercetin-3-O- -L-rhamnopyranosidewere also reported in the plant (Haque et al 2014).

The study revealed that the phenolic compounds and flavonoids in the ethanolic extract of leaves of *D. falcata* are responsible for the hepatoprotective activity of both ethanol and aqueous extracts significantly reduced AST, ALT, ALP, TB levels and increased the total protein (TP) and total albumin (TA) levels. The liver histology of the ethanolic extract treated group showed microfatty changes with a dense collection of lymphoid cells suggesting evidence of very little necrosis or degeneration (Pattanayak and Priyashree 2008).

Haque et al reported ethanolic and aqueous extract shows hepatoprotective activity against CCl₄ toxicity in rats indicated by a reduction AST, ALT, ALP, TP and TB (Haque et al 2014).

Ficus carica

Ficus carica L. (Family: Moraceae) is ordinarily deciduous tree and commonly known as "fig" and inhabited to southwest Asia and the eastern Mediterranean. The dried fruits of *F. carica* have been reported as source of vitamins, minerals, carbohydrates, sugars, fiber, organic acids, and phenolic compounds such as proanthocyanidins. Its fruit, root, and leaves are used in traditional medicine to treat various ailments such as gastrointestinal (i.e. colic, indigestion, loss of appetite and diarrhea), respiratory (i.e. sore throats, coughs, and bronchial problems), and cardiovascular disorders and as anti-inflammatory and antispasmodic remedy (Mawa et al. 2013).

Phytochemical studies on *F. carica* revealed the presence of various bioactive compounds such as phenolic compounds, phytosterols, organic acids, anthocyanin, triterpenoids, coumarins, and volatile compounds such as hydrocarbons, aliphatic alcohols (Oliveira et al. 2009).

The ethanolic extract of *F. carica* leaf (200 mg/kg) exhibited hepatoprotective activity against CCl₄ induced hepatotoxicity in mice model and observed that enhanced protection against CCl₄ induced hepatic damage (Aghel et al. 2011).

Petroleum ether extracts of Shade dried leaves of *Ficus carica* indicating promising antihepatotoxic activity against rifampicin induced liver toxicity in rats and there was significant reversal of biochemical, histological and functional changes i.e. SGOT,

SGPT, TB and histological changes in liver (Gond and Khadabadi 2008).

The ethanolic extract of *F. carica* leaves exhibited hepatoprotective and antioxidant activity in hepatotoxic Albino rats induced via CCl₄ and significantly decline biochemical parameters such as SGOT, SGPT, TP, TA, ALP and TB. The extract in addition reduced CCl₄ induced lipid peroxidation in-vivo and in-vitro (Mohd).

Hibiscus rosasinensis

Hibiscus rosasinensis flower extract is commonly used to treat symptoms related to blood circulation deficiencies, and is well known to reduce blood and plasma viscosity and thus improve microcirculation. In addition, it has received much attention due to its numerous biological activities, such as inhibition of platelet aggregation, suppression of hypertension and anti-aging (Biswas et al. 2014).

Phytochemical analysis of the crude extract of *H. rosasinensis* showed the presence of alkaloids, saponins, tannins, phenolics and flavonoids. Flower extract of *H. rosasinensis* exhibits hepatoprotective potential against hypercholesterolaemic rats and significantly decreased the levels of AST, ALT, ALP enzymes in the serum and significantly increased in TP levels (Biswas et al. 2014).

Hibiscus sabdariffa

Hibiscus sabdariffa L., also known as roselle, is an ideal crop for developing countries and

can be used as food and fibre. In China the seeds are used for their oil and the plant is used for its medicinal properties, while in West Africa the leaves and powdered seeds are used in meals. Additionally, it is used in the pharmaceutical and food industries (Da-Costa-Rocha et al. 2014).

The main phytochemicals of *H. sabdariffa* are organic acids, anthocyanins, polysaccharides and flavonoids. Methanolic and aqueous extracts of *H. sabdariffa* (100-800 mg/Kg) showed hepatoprotective effects in a range of models based on toxin-induced hepatitis including, *tert*-butylhydroperoxide, lipopolysaccharides, azathioprine, carbon tetrachloride, cadmium, ammonium chloride, acetaminophen and irradiation (Da-Costa-Rocha et al. 2014).

The hepatoprotective and antioxidant potential investigation showed that Polyphenol Rich Extract of *H. sabdariffa* (50 and 100 mg/kg) was able to scavenge the ABTS and DPPH radicals and significantly decline serum ALT, AST, ALP, LDH GSH, SOD, CAT and TBARS levels against CCl₄ induced liver toxicity in rats (Adetutu and Owoade 2013).

Leptadenia pyrotechnica

Leptadenia pyrotechnica (Family: Asclepiadaceae) is a plant wild growing in the Sharm El-Sheikh region, southern Sinai, Egypt. The leaves and bark of the plant are used in folk medicine to prepare antispasmodic, anti-inflammatory, antihistaminic, antibacterial diuretic, urolith

expulsion, expectorant, gout, and rheumatism remedies (Amal et al. 2009).

The main phycoconstituents of *L. pyrotechnica* are phenolic compounds, flavonoids, quercetin-3-*O*-galactoside, alkaloids, pregnane glycosides, amino acids, sterols, sitosterol, triterpenoids, taraxerol, fernenol, and leptadenol, fatty acids, and fatty alcohols (Amal et al. 2009).

The methanolic extract of whole plant of *Leptadenia pyrotechnica* exhibited hepatoprotective activity against hepatic damage in rats and marked reduction in the elevated activities of the hepatic enzymes i.e. SGOT, SGPT, ALP and TB levels (Partap et al. 2014).

Loranthus parasiticus

Loranthus parasiticus (Family: Loranthaceae) is aerial hemiparasitic plants. L. parasiticus contains pronounced levels of crude 82.28% of fiber, 2.70% of crude protein, 0.77% of crude fat and 2.60% of other components (Chen et al., 2003). Coriaria lactone is a mixture of compounds isolated from L. parasiticus includes sesquiterpene lactones such as coriamyrtin, tutin, corianin and coriatin. These sesquiterpenes were isolated from the ethanol extract of L. parasiticus leaves. Aqueous fraction of L. parasiticus contains two known proanthocyanidins of AC trimer and ()-catechin (Wong et al., 2012b).

The ethanol extract of *L. parasiticus* stem shows hepatoprotective potential against CCl₄

damage in rat liver and caused 50% inhibition on SGPT (Moghadamtousi et al. 2014).

Melastoma malabathricum

Melastoma malabathricum L. (family: Melastomaceae) is known to the "Senduduk" and has been used to treat various diseases. The leaves of M. malabathricum have been proven to possess various pharmacological activities, including antioxidant activity, antibacterial. antiviral, antiparasitic, cytotoxicity, anticoagulant, platelet-activating factor inhibitory, wound healing, antiulcer, antidiarrheal, antivenom, antiinflammatory, antinociceptive and antipyretic and hepatoprotective activity (Yahya et al. 2012).

Various phytochemical have been isolated from ethanolic extracts of M. malabathricum i.e. isoquercitrin 6"-O-gallate, malabathrin- A, -B, -C, -D, -E and -F, 1,4,6-tri-O-galloyl- -Dglucoside, 1,2,4,6-tetra-O-galloyl- -Dglucoside, strictinin, casuarictin, pedunculagin, nobotanin-B, -D, -G, -H and -J, pterocarinin C, new complex tannins in which an ellagitannin and a flavan-3-ol are bound by a C-glycosidic linkage belonging to type II + tannins, casuarinin, (–)-epicatechin gallate, epicatechin, stachyurin, procyanidin-B2 and -B5, stenophyllanins A and B, alienanin B, and brevifolincarboxylic acid. The methanol extract of M. malabathricum contains ursolic acid, 2-hydroxyursolic acid and asiatic acid, as glycerol-1,2-dilinolenylwell 3-0- as Dgalactopyranoside 1.2and glycerol dilinolenyl- 3-O- (4,6-di-O-isopropylidene)- -D-galactopyranoside, 2,5,6-trihydroxynaphtoic

carbonic acid. methyl-2,5,6trihydroxynaphtalene carbonate, flavonol glycoside derivative quercitrin and kaempferol-3-O-(2',6'-di-O-p-transcoumaroyl)- -glucosid. The hexane fraction of methanol extract M. malabathricum contains -sitosterol, -amyrin, uvaol, quercetin, quercitrin, rutin, and sitosterol-3-O--D-glucopyranoside; iv) 90% aqueous methanolic extract - ursolic acid, 2 hydroxyursolic acid and asiatic acid, sitosterol 3- O- -Dglucopyranoside, glycerol 1,2-dilinolenyl-3-O- -D-galactopyranoside and glycerol 1,2-dilinolenyl-3-O-(4,6-Oisopropylidene)- - D-galactopyranoside. The ethyl acetate extract M. malabathricum contains 2,5,6-trihydroxynaphtoic carbonic acid, methyl-2,5,6-trihydroxynaphtalene carbonate, and flavonol glycoside derivative, quercetin and quercitrin. The hexane extract M. malabathricum contains 2.5.6trihydroxynaphtoic carbonic acid, methyl-2,5,6-trihydroxynaphtalene carbonate, and flavonol glycoside derivative, -amyrin, patriscabatrine and auranamide (Mamat et al. 2013).

Mamat et al. reported hepatoprotective activity of the methanolic extract of *M. malabathricum* against the paracetamol induced hepatotoxic model in rats (Mamat et al. 2013)

Oxalis corniculata

Oxalis corniculata L., (Family: Oxalidaceae) is a subtropical, delicate-appearing, low growing, herbaceous plant being native of India abundantly distributed in damp shady places, roadsides, plantations, lawns, nearly all regions throughout the warmer parts of India, especially in the Himalayas and commonly known as creeping woodsorrel. Traditionally the plant reported to possess varsatile medicinal uses likely treatment for relieve the intoxication produced by *Datura*, as a refrigerant. decoction of roots is useful for worms, giddiness, diarrhea and dysentery. The leaves *O. corniculata* are useful for cough, cold, fever, antihelmintic, stomach ache and stop bleeding from wounds.

Phytochemical investigations methanolic and ethanolic extracts of of *O. corniculata* have revealed the presence of tannins, palmitic acid, fiber, tannin oleic, linoleic, linolenic, stearic acids, carbohydrate, glycosides, phytosterols, phenolic compounds, flavanoids, proteins, amino acids volatile oil, tartaric acid, citric acids, calcium oxalate and flavones (Das et al. 2012).

The ethanolic extract of *O. comiculata* exhibited significant hepatoprotective effects against the paracetamol induced hepatotoxicity in rat model and decline the serum levels of various biochemical parameters such as SGOT, SGPT and ALP (Sreejith et al. 2014).

Petroselinum crispum

Petroselinum crispum (family: Umbelliferae), locally known as Baqdunis and have been employed in the food, pharmaceutical, perfume, and cosmetics industries and serve as vegetable or condiment in cookery. Parsley has been claimed in Arab Traditional Medicine to

possess variety of properties including laxative, diuretic and antiurolithiatic. The leaves are used against inflammatory condition, aphrodisiac, mastitis, haematomata, constipation, flatulence, jaundice, edema, rheumatism, diseases of prostate and ailments, antimicrobial, antianemic, hemorrhagic, anticoagulant, antihyperlipidemic, antihep-atotoxic lumbago, blood pressure regulator, eczema, knee, ache, impotence nose bleed and laxative. Parsley seed are also used as a diuretic and the hypoglycemic activity. Jassim reported antihepatotoxic potential and protective activity on kidney damage induced by acid (Jassim 2013). valoproic The Phytochemical screening P. crispum revealed the presence of ascorbic acid, carotenoids, coumarins, apiole, various terpenoic compounds, phenyl propanoids, phthalides, furano coumarins, and tocopherol, flavonoids, tannins, sterols and or triterpenes (Al-Howiriny et al. 2003, Jassim 2013).

An ethanolic extract of *P. crispum* exhibited significant potential of anti-inflammatory and anti-hepatotoxic activities against inflammation induced by carrageenan and cotton pellet granuloma and hepatic damage induced by CCl₄ in rats. (Al-Howiriny et al. 2003).

Farzaei et al., reported hepatoprotective activity of ethanolic extract of *P. crispum* against CCl₄ induced oxidative stress mice (Farzaei et al. 2013).

Rheum palmatum

Rheum palmatum found among the oldest and best known herbal medicines. They are present in Chinese Pharmacopeia and used as laxative, antiphlogistic and treatment of indigestion. R. palmatum herb is also applied to stimulate gastric juice and bile excretion. Rhubarb roots have an antibacterial, anticancer activity, antioxidant activity, antiviral and hepatoprotective. The most important phytoconstituents of roots are antrachinons (2– 8%) such as chrysophanic acid, emodin, aloeemodin, rhein and physcion, with their Oglycosides such asglucorhein, chrysophanein, glucoemodin; sennosides AE, reidin C and others, their glycosides, bianthrones, hydrolysable and condensed tannins (5-10%), mainly catechin, stilbenes (resveratrol, rhapontigenin), volatile oil (containing di-isobutyl paeonol, phaltate, cinnamic aldehyde, methyl eugenol); rutin, fatty acids, calcium oxalate and phenolic constituent such as flavonoids, stilbenes and anthraquinones (Akinpelu et al. 2010).

In addition to the clinical evidence of the effectiveness of rhubarb in treating chronic hepatic diseases, many experiments at the animal, cellular and molecular levels have revealed that rhubarb and its anthraquinone constituents exert hepatoprotective effect against hepatic injury against CCl₄ induced elevation of ALT, AST, hyaluronic acid (HA) and laminin (LN) levels in rats and mice (Wang et al 2011).

Salvia miltiorrhiza

Salvia miltiorrhiza, (Family: Labiatae) is a deciduous perennial flowering plant in the genus Salvia, highly valued for its roots in traditional Chinese medicine and commonly known as Danshen and found in China and Japan, preferring grassy places in forests, hillsides, and along stream banks in the west and southwest provinces of China. Danshen is the dried root of S. miltiorrhiza and is one of the most versatile Chinese herbal drugs. Danshen was used infrequently in ancient Chinese's medicine, yet it has become an important herb in modern Chinese clinical practice and used clinically to treat and prevent cardio-vascular disease, hyperlipidemia, and cerebro-cardiac, vascular disorders such as atherosclerosis (Cheng 2007).

The main phyto-constituents of S. miltiorrhiza can be divided into two groups, hydrophilic compounds such as salvianolic acids, and lipophilic chemicals, including diterpenoid and tanshinones and the second group of components, labelled tanshinone I, tanshinone II, cryptotanshinone. More recently, nearly 40 variants of the basic tanshinone structures have been found in the roots. The tanshinones are unique chemical constituents, and similar compounds are not found in other Chinese herbs. The total tanshinone content of the roots is about 1%, with tanshinone I and II and cryptotanshinone being present in the largest amount. In one recent study, the concentration of S. miltiorrhiza yielded tanshinone II 0.29%, cryptotanshinone 0.23% and tanshinone I 0.11%. Among the tanshinones, tanshinone I, tanshinone IIA and cryptotanshinone are the major bioactive constituents and have various

kinds of pharmacological effects including anti-inflammatory, antibacterial, hepato-protective, antioxidant, antitumor activities, prevention of angina pectoris and myocardial infarction (Wang 2010).

S. miltiorrhiza has been widely used for the treatment of various liver diseases in different experimental models. Parajuli et al. reported hepatoprotective potential of purified extract isolated from S. miltiorrhiza enriched with tanshinone I, tanshinone IIA and cryptotanshinone on hepatocyte injury induced by CCl₄ in vitro and in vivo (Parajuli et al. 2013).

Tephrosia purpurea

Tehrosia purpurea (Family: Fabaceae) is common known as wild indigo in Tamil 'Kolanji' and widely distributed in tropical, sub-tropical and arid regions of the world. It is an important component of some preparations such as Tephroli and Yakrifit used for liver disorders. Traditionally drug is used as liver tonic, treatment of jaundice, dyspepsia, diarrhoea, rheumatism, asthma and urinary disorders (Dalwadi et al. 2014).

The crystalline compounds were isolated from petrol soluble fraction of CHCI₃ extract of *T. purpurea* along with isolonchocarpin, pongamol, lanceolatin B and lanceolatin A, isopentenyl. Phytochemical investigation also revealed the presence of flavonoids, glycosides, rotenoids, isoflavones, flavanones, chalcones, flavanols and sterols. (Dalwadi et al. 2014).

The ethanolic extract of the root of *T. purpurea* exhibits hepatoprotective activity against CCl₄ induced oxidative damage and dysfunction in the liver of rats and decline hepatospecific enzymes such as AST, ALT, acid phosphatase (ACP), ALP, LDH and 5' nucleotidase (5'NT) and serum levels of TB and total cholesterol (TC) whereas TP, TA, total glycogen (TG) and hepatospecific enzymes were significantly increased (Dalwadi et al. 2014).

Gora and Baxla reported hepatoprotective potential of *T. purpurea* extracts (500 mg/kg) against arsenic induced toxicity in wistar albino rats and observed reduced serum ALT, AST, ALP activity and increased TP and reduced necrosis and inflammation in liver (Gora and Baxla).

Aerial parts of *T. purpurea* evaluated for its efficacy in rats by inducing hepatotoxicity with D-galactosamine HCl (acute) and carbon tetrachloride (chronic). The ethanolic extract of *T. purpurea* leaves were evaluated for hepatoprotective activity in rats by inducing hepatotoxicity with CCl₄ and conclude that the hepatoprotective activity was more in ethanolic extract of leaves (Chaudhari et al. 2012).

Terminalia arjuna

Terminalia arjuna (Family: Combretaceae) is a deciduous and ever green tree, standing 20-30m above ground level and found in Uttar Pradesh, South Bihar, Madhya Pradesh, Delhi and Deccan region near ponds and livers. Stem bark *T. arjuna* possesses glycosides, large

quantities of flavonoids, tannins and minerals. Flavonoids have been detected to antioxidant, anti inflammatory and lipid lowering effects while glycosides are cardiotonic (Doorika and Ananthi 2012).

The aqueous extract of *T. arjuna* (200mg/kg) bark exhibits hepatoprotective effect against isoniazid induced acute liver damage on albino rats and significantly reduced the elevated levels of biochemical markers i.e. ALT, AST and ALP and increased the level of SOD and GSH (Doorika and Ananthi 2012).

The extract of bark of *Terminalia arjuna* reported for its hepatoprotective and antioxidative effects on cadmium induced toxicity in albino rats and significantly decline the serum levels of following biomarkers ALT, AST, ALP, and MDA and enhance the protein and antioxidant enzymes i.e. SOD, CAT, and GSH (Haidry and Malik 2014).

Trigonella foenum-graecum

Trigonella foenum-graecum L. (Family: leguminosae) is commonly known fenugreek. T. foenum-graecum with common name Methi in India is used in medicine to tonify kidneys, disperse cold and alleviate pain, hernia and pain (Sharma et al., 1996). T. foenum-graecum have therapeutic effect against anti ulcer, wound healing, CNS stimulant, immunomodulatory, antioxidant, antidiabetic, anti-neoplastic, anti-inflammatory and anti-pyretic drugs (Zargar 2014).

The extract prepared from the dried seeds of *T*. *foenum-graecum* exhibits hepatoprotective

potential against rat model induced liver cirrhosis by thioacetamide. After the administration of extract the oxidative stress and lipid peroxidation were reversed and the elevated levels of ALP, c-glutamyl transferase and selected biochemical markers of liver cirrhosis including drug metabolizing enzymes were also reversed (Zargar 2014).

Ziziphus mucronata

Ziziphus mucronata (Family: Ramnaceae) medicinal plants are used traditionally to treat various ailments in Africa and grows throughout Upper Egypt, Sinai and in the Mediterranean region, Africa, Australia, and tropical America.. Zizyphus has a common name "Nabka". The roots, bark and leaves are used in the treatment of arthritis, chest pains digestive disorders, weakness, liver complaints, obesity, urinary troubles, diabetes, skin infections, loss of appetite, fever, pharyngitis, bronchitis, anemia, diarrhea, and insomnia (Koeven 2001). Other medicinal properties of Z. mucronata are hypoglycemic, hypotensive anti-inflammatory, antimicrobial, antioxidant, antitumour and Hepatoprotective, antioxidant and as an immune system stimulant (Yossef et al. 2011).

Phytochemical studies show the presence of cyclopeptide alkaloids, flavonoids, sterols, tannins, and triterpenoid saponins and peptide, cyclopeptide betulinic acid (Yossef et al. 2011).

The methanol leaf extract of *Z. mucronatai* exhibits the hepatoprotective activity against dimethoate induced liver toxicity and oxidative stress in male albino Sprague Dawley rats by maintaining normal reduced GSH, vitamin C and E, SOD, CAT, cholineasterase and lipid profiles and significant decline activities of SGOT, SGPT and ALP (Kwape et al. 2013a).

Conclusion

The above mentioned herbal plants reviewed in detailed for hepatoprotective potential. In traditional systems of medicine herbal plants and their formulations are used to cure liver diseases. Despite the availability of extensive information concluded that herbal remedies are effective alternatives for management of hepatic diseases. A single drug cannot be effective against all types of liver disorders therefore effective formulations have to be developed using indigenous herbal plants, with proper pharmacological experiments and clinical trials.

Table: 1- List of harbal plants showing Hepatoprotective activity

S. N.	Name of plant	Plant Part used	Plant Extract	Active constituent	Experimental Model	Hepatoprotective activity/study outcome	Refere nce
1	Allium cepa	Fresh bulbs	Aqueous extract	Proteins, carbohydrates, polyphenolic compounds tannins, saponins, flavanoids	Ethanol induced liver damage in male rats	AST, ALT, ALP and TB, were significantly decreased	Kumar et al. 2013
2	Alocasia indica	Tuber	Ethanolic (80%) and Aqueous extract	Alkaloids, flavonoids, glycosides, saponin and tannins	CCl ₄ induced hepatic injuries in male Albino Wister rats	Recovery percentage of serum ALT by 65.32% and AST by 77.36%	Pal et al. 2014
3	Antrodia Cinnamomea	Fruiting bodies and Mycelia	Aqueous extract and Ethanolic extract (90%)	Benzenoids, diterpenes, triterpenoids, steroids, maleic/succinic acid derivatives,	CCl ₄ induced liver injury and ethanol induced liver injury in rats	Suppression of ethanol and CCl ₄ induced elevation of expression of hepatic mRNAs, i.e. MMP-9, TNF-, KLF-6, and TGF-1 levels	Liu et al. 2012
4	Bidens pilosa	Dried aerial parts	Ethanolic extracts (90%)	Flavonoids and polyacetylenes	CCl₄ induced liver injury in Male Balbc mice.	Significant decrease serum enzymatic activities of AST, ALT, and LDH	Kvieci nski et al. 2011
5	Boerhavia diffusa	Roots	ethanolic extract (95%)	Flavonoid glycosides, isoflavonoids, steroids, alkaloids, and phenolic and lignan glycosides	Hepatotoxicity induced by country made liquor in rats	Reduced the increment in serum parameters like SGPT, SAP, TGs, and total lipid levels	Mishra et al. 2014a
6	Caesalpenia crista	Leaves	Ethanolic extract (90%)	Carbohydrate, alkaloids, glycosides and phenolic compounds	Paracetamol induced hepatotoxicity in rats.	Reversed the levels of SGPT, SGOT, ALP, TB and TGs	Mishra et al. 2014
7	Chelidonium majus	Whole plant	Ethanolic extract	Benzyl isoquinoline alkaloids viz. protoberberine, protopine, benzophenanthredine	p-dimethyl aminoazobenzene (p-DAB) induced hepatocarcinogenesi s in mice	Biochemical assay of some toxicity marker enzymes and histology of liver sections suggest hepatoprotective effects	Biswas et al. 2008
8	Cyperus rotundus	Leaves	Methanolic extract	Flavonoids and alkaloids	CCl ₄ induced liver damage in wistar albino rats	Lowering the serum levels of various biochemical parameters such as serum SGOT, SGPT, ALP	Rao et al 2014
9	Dendrophtho e falcate	Leaves	Aqueous and Ethanolic extracts	Phenol and flavonoids	Liver damage was induced by intraperitonial administration of 25% CCl ₄ in olive oil in rats	Reduced the increment in serum parameters like AST, ALT, ALP, TP and TB	Haque et al. 2014
10	Ficus carica	Leaves, Fruit and Roots	Petroleum ether extract, Aqueous extract and Methanolic extract	Phenolics organic acids and volatile compounds	Rifampicin induced liver damage in male rats	Significant reversal of biochemical, histological, and functional changes induced by rifampicin on rats indicated potential hepatoprotective activity	Mawa et al. 2013
11	Hibiscus rosa sinensis	Flower	Aqueous extract	Saponins, flavonoids, tannins, phenols, sterols, alkaloids, and anthocyanins.	Induced hypercholesterolemi a by feeding pure cholesterol and cholic acid orally mixing with coconut oil in rats	Decreased the levels of AST, ALT, ALP enzymes in the serum	Biswas et al. 2014
12	Hibiscus sabdariffa	Seeds, Leaves and Shoots	Aqueous extracts	Phenolic acids, organic acid and anthocyanins	CCl ₄ induced oxidative damage of rat liver	Significantly reduced serum activities of ALT, AST, and ALP	Da- Costa- Rocha et al. 2014
13	Leptadenia pyrotechnica	Whole plant	Methanolic, petroleum ether, chloroform, acetone and aqueous extract	flavonoids and polyphenolic compounds	Paracetamol induced liver damage in wistar rats	A marked reduction in the elevated activities of the hepatic enzymes i.e. SGOT, SGPT, ALP and TB levels	Partap et al. 2014
14	Loranthus parasiticus	Leaves	Ethanolic extract	Sesquiterpene lactones (coriamyrtin, tutin, corianin and coriatin)	D-galactosamine and CCl ₄ damage in rat liver cells	50% inhibition on SGPT	Mogha damtou si et al. 2014

15	Melastoma malabathricu m	Leaves	Methanolic extract	Flavonoids	Paracetamol induced liver toxicity in rats	Serum liver enzymes ALT, ALP and AST as well as the microscopic observations and microscopic scoring supported the hepatoprotective potential	Mamat et al. 2013
16	Oxalis corniculata	Whole plants	Ethanolic extract (95%)	Flavonoids, phenols and tocopherols	Paracetamole induced hepatotoxicity in wistar rats	Lowering the serum levels of various biochemical parameters such as SGOT, SGPT and ALP,	Sreejith et al. 2014
17	Petroselinum crispum	Leaves	Aqueous extract	Flavonoids, phenolic compounds and ascorbic acid	STZ- induced diabetic rats	Significant decrease in blood glucose, ALP, sialic acid, uric acid, potassium and sodium levels, liver lipid peroxidation and non- enzymatic glycosylation and increase in liver glutathione	Farzaei et al. 2013
18	Rheum palmatum	Dried root	Ethanolic extract (90%)	Anthraquinone derivatives and tanninrelated compound	CCl ₄ induced liver injury in rats	Elevation of ALT, AST, HA and laminin (LN) levels were reversed	Wang et al 2011
19	Salvia miltiorrhiza	Dried pulverized roots	Ethanolic extract (95.6%)	Salvianolic acids, tanshinone, cryptotanshinone	CCl ₄ induced liver injury in rats	Induce apoptosis of hepatic stellate cells (HSCs)	Parajuli et al. 2013
20	Tephrosia purpurea	Root	Ethanolic extract	Isolonchocarpin, purpurenone, purpurin, dehydrolsodericin, maackiain, purpurin, semiglabrin and pseudosemiglabrin	CCl ₄ induced oxidative damage and resultant dysfunction in the liver of rats.	Decreasing the serum levels of AST, ALT, ACP, ALP, LDH and 5' NT	Dalwad i et al. 2014
21	Terminalia arjuna	Bark	Aqueous extract	Flavonoids, tannins and oligomeric proanthocyanidins	Cadmium induced toxicity in Albino Rats	Significantly reversed the elevated the serum levels of following biomarkers ALT, AST, ALP, and MDA	Haidry and Malik 2014
22	Trigonella foenum- graecum	Dried seeds	Ethanolic extract	-carotein, saponins, coumarin, nicotinic acid, phytic acid, scopolatin, trigonelline	Thioacetamide induced liver cirrhosis in rats	The elevated levels of alkaline phosphatase, c- glutamyl transferase and selected biochemical markers of liver cirrhosis were reversed.	Zargar 2014
23	Ziziphus mucronata	Leaves	Methanolic extract (70%)	Phenols	Dimethoate induced liver damage in rats	A significant decline serum marker enzymes SGOT, SGPT and ALP	Kwape et al. 2013

References

Adetutu A, Owoade AO. Hepatoprotective and Antioxidant Effect of *Hibiscus* Polyphenol Rich Extract (HPE) Against Carbon Tetrachloride (CCL4) –Induced Damage in Rats. British Journal of Medicine & Medical Research 2013; 3:1574-1586.

Aghel N, Kalantari H, Rezazadeh S. Hepatoprotective Effect of *Ficus carica* Leaf Extract on Mice Intoxicated with Carbon Tetrachloride. Iranian Journal of Pharmaceutical Research 2011; 10:63-68.

Akinpelu DA, Aiyegoro OA, Okoh AI. The in vitro antioxidant property of methanolic extract of *Afzelia Africana* (Smith.). J Med Plant Res 2010; 4:2022-2027.

Alcaraz M J, Jimenez MJ. Flavonoids as antiinflammatory agents. Fitoterapia 1998; 59:25– 38.

Al-Howiriny TA, Al-Sohaibani MO, El-Tahir KH, Rafatullah S. Preliminary evaluation of the anti-inflammatory and anti-hepatotoxic activities of 'Parsley' *Petroselinum crispum* in rats. Journal of Natural Remedies 2003; 3:54–62.

Amal M. Moustafa Y, Khodair AI, Saleh M A. Isolation, structural elucidation of flavonoid constituents from *Leptadenia pyrotechnica* and evaluation of their toxicity and antitumor activity. Pharmaceutical Biology 2009; 47:539–552.

Ao ZH, Xu ZH, Lu ZM, Xu HY, Zhang XM, Dou WF. Niuchangchih (*Antrodia camphorata*) and its potential in treating liver diseases. J Ethnopharmacol 2009; 121:194-212.

Bairwa K, Srivastava A, Jachak SM. Quantitative analysis of Boeravinones in the roots of *Boerhaavia diffusa* by UPLC/PDA. Phytochemical Analysis 2014; 25:415-420.

Bartolome AP, Villasenor IM, Yang WC: *Bidens pilosa* L. (Asteraceae). Botanical Properties, Traditional Uses, Phytochemistry, and Pharmacology. Evidence-Based Complementary and Alternative Medicine 2013: 2013.

Biswas A, D'Souza UJA, Bhat S, Damodar D. The hepatoprotective effect of *Hibiscus rosa sinensis* flower extract on diet-induced hypercholesterolemia in male albino wister rats. Int J Med Pharm Sci 2014; 4.

Biswas J, Bhattacharjee N, Khuda-Bukhsh AR. Efficacy of a plant extract (*Chelidonium majus* L.) in combating induced hepatocarcinogenesis in mice. Food Chem Toxicol 2008; 46:1474–1487.

Bott RF, Labuza TP, Oliveira WP. Stability testing of spray- and spouted bed-dried extracts of *Passiflora alata*. Dry Techno 2010; 28:1255–1265.

Chaudhari TB, Tambe DA, Chaudhari S R. Phytopharmacology of *Tephrosia purpurea* Pers. (Fabaceae) - A Review. Journal of Pharmacognosy and Herbal Formulations 2012; 2:8.

Chen W, Weng YM, Tseng CY. Antioxidative and antimutagenic activities of healthy herbal drinks from Chinese medicinal herbs. Am J Chin Med 2003; 31:523–532.

Cheng TO. Cardiovascular effects of *Danshen*. J Cardiol 2007: 121:9–22.

Chiang PC, Lin SC, Pan SL, Kuo CH, Tsai IL, Kuo MT, et al. Antroquinonol displays anticancer potential against human hepatocellualr carcinoma cells: a crucial role of AMPK and mTOR pathway. Biochem Pharmacol 2010; 79:162-171.

Da-Costa-Rocha I, Bonnlaender B, Sievers H, Pischel I, Heinrich M. *Hibiscus sabdariffa* L. - a phytochemical and pharmacological. Food Chemistry 2014; 165:424–443.

Dalwadi PP, Patel JL, Patani PV. *Tephrosia purpurea* Linn (Sharpunkha, Wild Indigo): A Review on Phytochemistry and Pharmacological Studies. Indian J Pharm Biol Res 2014; 2:108-121.

Das K, Kathiriya AK, Kumar E P, Benson MK, Einstein JW. Evaluation of hepatoprotective activity of aqueous and ethanolic extract of *Oxalis corniculata* against intoxication of thioacetamide induced rats. Revista Brasileira de Farmacognosia Brazilian Journal of Pharmacognosy 2012; 22:412-417.

Dominiczak M H. Lipids and Lipo propteins. In: Baynes JW, Dominiczak MH (Eds) Medical Biochemistry, 2005, (pp 234- 242.). Elsevier mosby. Philadephia.

Doorika P, Ananthi T. Antioxidant and Hepatoprotective properties of *Terminalia arjuna* Bark on Isoniazid Induced Toxicity in Albino rats. Asian J Pharm Tech 2012; 2:15-18.

Dorant E, Brandt PAV, Goldbohm RA. A prospective cohort study on the relationship between onion and leek consumption, garlic supplement use and the risk of colorectal

carcinoma in The Netherlands. Carcinogenesis 1996; 17:477–484.

Elhassaneen YA, Sanad MI. Phenolics, selenium, vitamin C, amino acids and pungency levels and antioxidant activities of two Egyptian onion varieties. Am J Food Technol 2009; 4:241–254.

Fang Z, Bhandari B. Effect of spray drying and storage on the stability of bayberry polyphenols. Food Chem 2011; 129:1139–1147.

Farzaei MH, Abbasabadi Z, Ardekani MRS, Rahimi R, Farzaei F. Parsley: a review of ethnopharmacology, phytochemistry and biological activities. J Tradit Chin Med 2013; 33: 815-826.

Gilcaa M, Gamana L, Stoiana EPI, Atanasiua V. *Chelidonium majus* – an Integrative Review: Traditional Knowledge versus Modern Findings. Forsch Komplementmed 2010; 17:241–248.

Gond NY, Khadabadi SS. Hepatoprotective activity of *Ficus carica* leaf extract on rifampicin-induced hepatic damage in rats. Indian Journal of Pharmaceutical Sciences 2008; 70:364–366.

Gora RH, Baxla SL, Roy BK. Hepatoprotective activity of *Tephrosia purpurea* against arsenic induced toxicity in rats, Indian Journal of Pharmacology 2014; 46:197-200.

Griffiths G L, Crowther TT. Onions – a global benefit to health. Phytother Res 2002; 16:603–615.

Gupta AK, Tandon N, Sharma M. Quality standards of Indian medicinal plants. Indian Council of Medical Research 2005; 29:25-26.

Haidry M, Malik A T. Hepatoprotective and Antioxidative Effects of *Terminalia Arjuna* against Cadmium Provoked Toxicity in Albino Rats (*Ratus Norvigicus*). Biochem Pharmacol 2014; 3:1.

Haque A, Tahmina, Afsana SK, Sarker IR, Hossain M, Islam S, et al. Antioxidant and hepatoprotective effects of aqueous and ethanol extracts of *Dendrophthoe falcate* Linn leaves. Archives 2014: 1:90-101.

Hseu YC, Chen SC, Yech YJ, Wang L, Yang HL. Antioxidant activity of *Antrodia camphorata* on free radicalinduced radicalinduced endothelial cell damage. J Ethnopharmacol 2008; 118:237-245.

Hsiao G, Shen MY, Lin KH, Lan MH, Wu LY, Chou DS, et al. Antioxidative and hepatoprotective effects of *Antrodia camphorata* extract. J Agric Food Chem 2003; 51:3302-3308.

Huang CH, Chang YY, Liu CW, Kang WY, Lin YL, Chang HC, et al. Fruiting body of Niuchangchih (*Antrodia camphorata*) protects livers against chronic alcohol consumption damage. J Agric Food Chem 2010; 58:3859-3866.

Ige SF, Akhigbe RE, Adewale AA. Effect of Allium cepa (onion) extract on cadmium-induced nephrotoxicity in rats. Kidney Res J 2011; 1:41–47.

Jassim AM. Protective Effect of *Petroselinum crispum* (parsley) extract on histopathological changes in liver, kidney and pancreas induced by Sodium Valproate- In male Rats. Kufa Journal For Veterinary Medical Sciences 2013; 4.

Jeong SJ, Miyamoto T, Inagaki M, Kim YC, Rotundines HR. Three novel sesquiterpene alkaloids from *Cyperus rotundus*. J Nat Prod 2000; 63:673-675.

Kannur DM, Hukkeri VI, Akki KS. Adaptogenic activity of *Caesalpinia bonduc* seed extracts in rats. J Ethnopharmacol 2006; 108:327-331.

Kilani S, Ledauphin J, Bouhlel I, Sghaier MB, Boubaker J, Skandrani I, et al. Comparative study of *Cyperus rotundus* essential oil by a modified GC/MS analysis method. Evaluation of its antioxidant, cytotoxic, and apoptotic effects. Chem Biodivers 2008; 5:729-742.

Koeven E. Medicinal, poisonous, and edible plants in Namibia. Windhoek (Namibia): Klause Hess Publishers 2001; pp335.

Kook S, Kim GH, Choi K. The antidiabetic effect of onion and garlic in experimental diabetic rats: meta-analysis. J Med Food 2009; 12:552–556.

Kumar KE, Harsha KN, Sudheer V, babu NG, In vitro antioxidant activity and in vivo hepatoprotective activity of aqueous extract of *Allium cepa* bulb in ethanol induced liver damage in Wistar rats. Food Science and Human Wellness 2013; 2:132–138.

Kumar KJ, Chu FH, Hsieh HW, Liao JW, Li WH, Lin JC, et al. Antroquinonol from ethanolic extract of mycelium of *Antrodia Cinnamomea* protects hepatic cells from ethanol-induced oxidative stress through Nrf-2 activation. J Ethnopharmacol 2011; 136:168-177.

Kumar S V S, Mishra H. Hepatoprotective Activity Of Rhizomes Of *Cyperus Rotundus* Linn Against Carbon Tetrachloride-Induced Hepatotoxicity. Pharmaceutical sciences 2005; 67:84-88.

Kviecinski MR, Felipe KB, Correia JFG, Ferreira EA, Rossi MH. Brazilian *Bidens pilosa* Linne' yields fraction containing quercetin-derived flavonoid with free radical scavenger activity and hepatoprotective effects. Libyan J Med 2011; 6:5651.

Kwape TE, Chaturvedi P, Kamau JM, George S. Hepato-protective potential of methanol extract of leaf of *Ziziphus mucronata* (zmlm) against dimethoate toxicity: biochemical and

histological approach. Ghana medical journal 2013; 47:112-120.

Kwape TE, Chaturvedi P, Kamau M, Majinda R. Anti-Oxidant and Hepatoprotective Activities of *Ziziphus mucronata* Fruit Extract Against Dimethoate-Induced Toxicity. Journal of Pharmacopuncture 2013; 16:21-29.

Liu YW, Lu KH, Ho CT, Sheen LY. Protective Effects of *Antrodia Cinnamomea* Against Liver Injury. Journal of Traditional and Complementary Medicine 2012; 2:284-294.

Lu Y, Cederbaum A I. CYP2E1 and oxidative liver injury by alcohol, Free Radic. Biol Med 2008; 44:723–738.

Lu ZM, Tao WY, Xu HY, Ao ZH, Zhang XM, Xu ZH. Further studies on the hepatoprotective effect of *Antrodia camphorata* in submerged culture on ethanol-induced acute liver injury in rats. Nat Prod Res 2011; 25:684-695.

Lu ZM, Tao WY, Zou XL, Fu HZ, Ao ZH. Protective effects of mycelia of *Antrodia camphorata* and *Armillariella tabescens* in submerged culture against ethanol-induced hepatic toxicity in rats. J Ethnopharmacol 2007; 110:160-164.

Mamat SS, Kamarolzaman MFF, Yahya F, Mahmood ND, Shahril MS. Methanol extract of *Melastoma malabathricum* leaves exerted antioxidant and liver protective activity in rats. BMC Complementary and Alternative Medicine 2013; 13:326.

Mawa S, Husain K, Jantan I. *Ficus carica* L. (Moraceae) Phytochemistry, Traditional Uses and Biological Activities. Evidence-Based Complementary and Alternative Medicine 2013; 2013.

Meagher EA, Barry OP, Burke A, Lucey MR, Lawson JA, Rokach J, et al. Alcohol-induced generation of lipid peroxidation products in humans. The J Clin Inv 1999; 104:805-813.

Miralles J, Noba K, Ba AT, Gaydou EM, Kornprobst JM. Chemotaxonomy in Nyctagynaceae family: sterols and fatty acids from the leaves of three *Boerhaavia* species. Biochemical Systematics and Ecology 1998; 16:475–478.

Mishra G, Khosa RT, Singh P, Jha KK. Hepatoprotective potential of ethanolic extract of *Caesalpenia crista* leaves against paracetamol induced hepatotoxicity in rats. Journal of Coastal Life Medicine 2014; 2:575-579.

Mishra S, Aeri V, Gaur PK, Jachak SM. Phytochemical, Therapeutic, and Ethnopharmacological Overview for a Traditionally Important Herb: *Boerhavia diffusa* Linn. BioMed Research International 2014; 2014;808302.

Mitra S, Sur RK, Roy A, Mukherjee AS. Effect of *Chelidonium majus* L. on experimental hepatic tissue injury. Phytother Res 1996; 10:354–356.

Moghadamtousi SZ, Kamarudin MNA, Chan CK, Goh BH, Kadir HA. Phytochemistry and Biology of *Loranthus parasiticus* Merr, a Commonly Used Herbal Medicine. The American Journal of Chinese Medicine 2014; 42:23–35.

Mujeeb M, Khan SA, Ali B. Hepatoprotective Activity of the Ethanolic Extract of *Ficus carica* Linn. LeavesinCarbon Tetrachloride-Induced Hepatotoxicityin Rats. Iran J Pharma Res 2011; 10:301-306.

Mulla WA, Chopade AR, Bhise SB, Burade KB, Khanwelkar CC. Evaluation of antidiarrheal and in vitro antiprotozoal activities of extracts of leaves of *Alocasia indica*. Pharm Biol 2011; 49:354-361.

Mulla WA, Kuchekar SB, Thorat VS, Chopade AR, Kuchekar BS. Antioxidant, Antinociceptive and Anti-inflammatory Activities of Ethanolic Extract of Leaves of

Alocasia indica (Schott.). J Young Pharm 2010; 2:137–143.

Mulla WA, Salunkhe VR, Kuchekar SB, Qureshi N. Free radical scavenging activity of leaves of *Alocasia indica* (Linn). Indian J Pharm Sci 2009; 71:303–307.

Obioha UE, Suru SM, Ola-Mudathir KF. Hepatoprotective potentials of onion and garlic extracts on cadmium-induced oxidative damage in rats. Biol Trace Elem Res 2009; 129:143–156.

Olaleye MT, Akinmoladun AC, Ogunboye AA, Akindahunsi AA. Antioxidant activity and hepatoprotective property of leaf extracts of *Boerhaavia diffusa* Linn against acetaminophen-induced liver damage in rats. Food and Chemical Toxicology 2010; 48:2200–2205.

Oliveira APP, Valentao JA, Pereira BM, Tavares SF, Andrade PB. *Ficus carica* L. metabolic and biological screening. Food and Chemical Toxicology 2009; 47:2841–2846.

Pal S, Bhattacharjee A, Mukherjee S, Bhattacharya K, Khowala S. Antioxidant and Hepatoprotective Activity of Ethanolic Extract of *Alocasia indica* Tuber. American Journal of Phytomedicine and Clinical Therapeutics 2014; 2:191-208.

Parajuli DR, Park EJ, Che XH, Jiang WY, Kim YC, Sohn DH, et al. PF2401-SF, Standardized Fraction of *Salvia miltiorrhiza*, Induces Apoptosis of Activated Hepatic Stellate Cells in Vitro and in Vivo. Molecules 2013; 18:2122-2134.

Park EJ, Zhao YZ, Kim YC, Sohn DH. Preventive effects of apurified extract isolated from *Salvia miltiorrhiza* enriched with tanshinone I, tanshinone IIA and cryptotanshinone on hepatocyte injury in vitro and in vivo. Food Chem Toxicol 2009; 47:2742–2748.

Partap S, Tewari U, Sharma K, Jha KK. Hepatoprotective activity of whole plant extract of *Leptadenia pyrotechnica* against paracetamol induced damage in rats. Journal of Drug Delivery & Therapeutics 2014; 4:36-39.

Pattanayak SP, Priyashree S. Hepatoprotective activity of the leaf extracts from *Dendrophthoe falcata* (L.f.) Ettingsh against carbon tetrachloride-induced toxicity in Wistar albino rats. Pharmacognosy magazine 2008; 4:218-222.

Pradhan SC, Girish C. Hepatoprotective herbal drug, silymarin from experimental pharmacology to clinical medicine. Indian J Med Res 2006; 124:491-504.

Price KR, Rhodes MJC. Analysis of the major flavonol glycosides present in four varieties of onion (*Allium cepa*) and changes in composition resulting from autolysis. J Sci Food Agric 1997; 74:331–339.

Ramaiah SK. A toxicologist guide to the diagnostic interpretation of hepatic biochemical parameters. Food Chem Toxicol 2007; 45:1551–1557.

Rao Y N, Rao MP, Rao VR, Suresh K, Rafi M, Rao TN, et al. Prelimeinary phytochemical screening and hepatoprotective activity of methanolic leaves extract of *Cyperus rotundus* in CCl₄ induced hepetotoxicity in albino wistar rats. World journal of pharmacy and pharmaceutical sciences 2014; 3:787-795.

Shahjahan M, Sabitha KE, Jainu M, Shyamala Devi CS. Effect of *Solanum trilobatum* against carbon tetrachloride induced hepatic damage in albino rats. Indian J Med Res 2004; 120:194-198.

Shi J, Aisaki K, Ikawa Y, Wake K. Evidence of Hepatocyte Apoptosis in Rat Liver after the Administration of Carbon Tetrachloride. Am J Pathol 1998; 153:515-525.

Silva FL, Fischer DCH, Tavares JF, Silva MS, De-Athayde-Filho PF, Barbosa-Filho JM. Compilation of secondary metabolites from *Bidens pilosa* L. Molecules 2011; 16:1070-1102.

Singh N, Kamath V, Narasimhamurthy K, Rajini PS. Protective effects of potato peel extract against carbonatetrachloride- induced liver injury in rats. Environ Toxicol Pharmacol 2008; 26:241-246.

Sivapalan SR. Medicinal uses and Pharmacological activities of *Cyperus rotundus* Linn –A Review. International Journal of Scientific and Research Publications 2013; 3.

Sreejith G, jayasree M, Latha PG, Suja SR, Shyamal S, Shine VJ. Hepatoprotective activity of *oxalis corniculata* L. ethanolic extract against paracetamole induced hepatoxicity in wistar rats and its in vitro antioxidant effects. Indian journal of experimental biology 2014; 52:147-152.

Teyssier C, Amiot MJ, Mondy N. Effect of onion consumption by rats on hepatic drugmetabolizing enzymes. Food Chem Toxicol 2001; 39:981–987.

Uddin SJ, Mondal K, Shilpi JA, Rahnan MT. Antidiarrhoeal activity of *Cyperus rotundus*. Fitoterapia 2006; 77:134.

Ujowundu CO, Igwe CU, Enemor VHA, Nwaoguand LA, Okafor OE. Nutritive and anti-nutritive properties of *Boerhavia diffusa* and *Commelina nudiflora* leaves. Pakistan Journal of Nutrition 2008; 7:90–92.

Venkatalakshmi P, Eazhisai VD, Netaji S. Hepatoprotective Activity of *Boerhavia diffusa* against paracetamol induced toxicity in rats. Journal of Chemical&Pharmaceutical Research 2011; 3:229–232.

Vulgaris T, Hossain MA, Salim KL, Raqmi AL, Mijizy ZH, Weli AM, et al. Study of total

phenol, flavonoids contents and phytochemical screening of various leaves crude extracts of locally grown. Asian Pac J Trop Biomed 2013; 3:705–710.

Wang BQ. *Salvia miltiorrhiza*: Chemical and pharmacological review of a medicinal plant. J Med Plant Res 2010; 4:2813-2820.

Wang JB, Zhao HP, Zhao YL, Jin C, Liu D J. Hepatotoxicity or Hepatoprotection? Pattern Recognition for the Paradoxical Effect of the Chinese Herb *Rheum palmatum* L. in Treating Rat Liver Injury. Plos One: Pattern Recognition for Herbal Toxicity 2011; 6:e24498.

Wong DZH, Kadir HA, Ling SK. Bioassay-guided isolation of neuroprotective compounds from *Loranthus parasiticus* against H2O2-induced oxidative damage in NG108-15 cells. J Ethnopharmacol 2012; 139:256–264.

Wu MF, Peng FC, Chen YL, Lee CS, Yang YY, Yeh MY, et al. Evaluation of genotoxicity of *Antrodia cinnamomea* in the Ames test and the in vitro chromosomal aberration test. In Vivo 2011; 25:419-423.

2685.

Yossef H E, Khedr A Aand Mahran M Z. Hepatoprotective activity and antioxidant effects of El Nabka (*Zizyphus spina-christi*) fruits on rats hepatotoxicity induced by carbon tetrachloride. Nature and Science 2011; 9.

Yuan LP, Chen FH, Ling L, Dou PF, Bo H, Zhong MM. Protective effects of total flavonoids of *Bidens pilosa* L. (TFB) on animal liver injury and liver fibrosis. J Ethnopharmacol 2008; 116:539-546.

Zargar S. Protective effect of *Trigonella* foenum-graecum on thioacetamide induced hepatotoxicity in rats. Saudi Journal of Biological Sciences 2014; 21:139–145.

Zielinska M, Gulden M, Seibert H. Effects of quercetin and quercetin-3- O-glycosides on oxidative damage in rat C6 glioma cells. Environ Toxicol Pharmacol 2008; 13:47–53.

Yahya F, Ismail NA, Kamisan FH, Mamat SS, Din SS, Zainulddin WWW, et al. Effect of aqueous extract of *Melastoma malabathricum* leaves against paracetamol- and CCl4-induced liver toxicity in rats. African Journal of Pharmacy and Pharmacology 2012; 6:2682-